

## Horizons of Psychiatric Genetics and Epigenetics: Where Are We and Where Are We Heading?

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Today multinational studies using genome-wide association scan (GWAS) for >1000,000 polymorphisms on >100,000 cases with major psychiatric diseases versus controls, combined with next-generation sequencing have found ~100 genetic polymorphisms associated with schizophrenia (SCZ), bipolar disorder (BD), autism, attention deficit and hyperactivity disorder (ADHD), etc. However, the effect size of each genetic mutation has been generally low (<1%), and altogether could portray a tiny fraction of these mental diseases. Furthermore, none of these polymorphisms was specific to disease phenotypes indicating that they are simply genetic risk factors rather than causal mutations.

The lack of identification of the major gene(s) in huge genetic studies increased the tendency for reexamining the roles of environmental factors in psychiatric and other complex diseases. However, this time at cellular/molecular levels mediated by epigenetic mechanisms that are heritable, but reversible while interacting with the environment. Now, gene-specific or whole-genome epigenetic analyses have introduced hundreds of aberrant epigenetic marks in the blood or brain of individuals with psychiatric diseases that include aberrations in DNA methylation, histone modifications and microRNA expression. Interestingly, most of the current psychiatric drugs such as valproate, lithium, antidepressants, antipsychotics and even electroconvulsive therapy (ECT) modulate epigenetic codes.

The existing data indicate that, the impacts of environment/nurture, including the uterine milieu and early-life events might be more significant than genetic/nature in most psychiatric diseases. The lack of significant results in large-scale genetic studies led to revise the bolded roles of genetics and now we are at the turning point of genomics for reconsidering environmental factors that through epigenetic mechanisms may impact the brain development/functions causing disease phenotypes.

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### Psychiatric Genetic: Accomplishments and Challenges

The advancement of genetic sciences and the development of new techniques greatly improved our understanding of mental diseases during the last decade and this is going to open new windows toward the diagnosis and treatment of psychiatric diseases in coming years. While ten years ago genetic studies were limited to single gene analysis in hundreds of cases and controls, today multinational consortiums have executed GWAS for >1000,000 known

polymorphisms (mutations) on hundreds of thousands of cases and controls affected by major psychiatric diseases. They found hundreds of genetic polymorphisms or copy number variations (CNVs) (due to deletion or insertion) associated with SCZ and BD, autism and ADHD, many of which were also identified by smaller traditional genetic association and family studies. Interestingly, several of these polymorphisms are shared by different mental diseases (1), particularly in SCZ and BD (2, 3). Some important shared genes include CACNA1C ( $\alpha$ -1C subunit of the L-type voltage-gated calcium channel), ZNF804A, PBRM1, neurogranin, SYNE1 and major histocompatibility region on chromosome 6 (4-7).

Despite this progress, the effect size of each genetic mutation (which most of them are intergenic), including CNVs and de novo

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mutations, has been generally low (<1%), and altogether could portray a tiny fraction of these mental diseases (3, 8). Some investigators suggest that, this small effect size could be due to the heterogeneity of mental diseases. Thus more sample sizes are required (e.g. >100,000). However, doubling the sample size during the last five years, combined with new analytical methods, could just increase the number of candidate genes, including those affected by de novo mutations, still with small effect size (9, 10). Notably, unlike known genetic diseases such as Huntington's Disease or thalassemia, none of these polymorphisms were specific to the disease phenotypes, including endophenotypes (11), and were only more frequent in disease state, compared to the controls. This indicates that these polymorphisms can be simply considered as risk factors, and there could not be ~100 types of SCZ or BD matching to ~100 identified genes. Hence, the actual cause(s) should be other players to be identified.

The GWAS findings in major depressive disorder (MDD) and obsessive-compulsive disorder (OCD) have been even more frustrating as no specific gene could surpass the threshold of significant association (12, 13), even in large studies with almost ten thousand cases of MDD versus controls (14). Although the development of next generation sequencing technology (the most advanced method for the detection of unknown mutations in whole exome/genome analysis) could help to identify a number of new candidate genes for SCZ (e.g. GRM5, PPEF2 and LRP1B), and autism (e.g. SCN2A), it has yet to introduce any gene(s) with a key role in major mental diseases (15-21).

While, these large-scale genetic studies have not been successful to tease out the main underlying genetic causes of major mental diseases, several pharmacogenomic studies support that specific genetic polymorphisms can predict the side effects and probability of responsiveness to specific psychiatric drugs (22-27). Hence, a number of companies have started to utilize a smaller set of genetic data to establish personalized treatment protocols for individuals affected by major mental

diseases (e.g. [www.genomind.com](http://www.genomind.com) and <http://genelex.com>). This new approach for personalized medicine is going to be complemented with the induced pluripotent stem (iPS) cell technology that can generate iPS cells from the peripheral cells of affected individuals containing all genetic mutations of the original cells. These cells can be differentiated to neuronal cells in culture (e.g. dopaminergic, serotonergic or GABAergic cells) and the effect of the candidate drugs can be evaluated in a Petri dish (28), like an antibiogram.

### **Psychiatric Epigenetics: Achievements and Opportunities**

The lack of identification of major gene(s) linked to psychiatric diseases in GWAS and deeper analyses using next generation sequencing technology, increased the tendency of scientific communities for reexamining the roles of environmental factors in impacting the brain development and functions. However, this time at cellular or molecular levels mediated by latterly discovered epigenetic mechanisms. Epigenetics refers to the science of complementary chromosomal codes (beside genetic codes) that govern cellular differentiations and regulate genes expression in a dynamic manner based on the tissue types, developmental periods and micro/macro-environmental conditions (such as hormonal effects, cell metabolic state, nutritional habits/status, seasonal or ecological conditions). So far, four major mechanisms have been introduced for epigenetic regulations, including DNA methylation, histone modifications, the interference of non-coding microRNAs and RNA editing (29).

Well-known varieties of DNA methylation include methylation of cytosine nucleotides that are followed by guanine or adenine, and hydroxymethylation that is a temporary product during the conversion of methylated cytosines to unmethylated cytosines. Unlike other tissues, hydroxymethylation is quite abundant in the human brain. While DNA methylation generally suppresses gene expression, hydroxymethylation can induce

gene expression and appears to play a key role in functional plasticity of neuronal cells that barely replicate. While several types of DNA methyltransferase enzymes induce DNA methylation, TET and IDH family of enzymes catalyze hydroxymethylation.

Histone modifications which comprise acetylation, methylation, ubiquitination, etc. of different amino acids of histone proteins may suppress or increase the expression of interconnected genes depend on the identity and location of those amino acids. Several types of enzymes such as histone acetylases, histone deacetylases (HDACs) (that is inhibited by valproate), histone methylases and demethylases are involved in histone modifications (30).

DNA methylation and histone modifications have complex interplays with more than 1000 recently discovered non-coding microRNAs (~20 bases in length) that can each target over 100 genes, even in other tissues (i.e., exosomal or circulating miRNAs). miRNAs generally suppress RNA expression or degrade the transcribed RNA, thus inhibit protein synthesis. RNA editing that is engaged in diverse splicing of the RNA transcripts (generating multiple isoforms of protein with functional diversities) makes epigenetic regulation even more complex.

During the last ten years, several epigenetic studies, using gene-specific or whole genome epigenetic analysis (e.g. the Illumina 27 k or 450 k DNA methylation array, and next generation sequencing following Immunoprecipitation of methylated DNA or acetylated/methylated chromatin) have introduced hundreds of aberrant epigenetic marks in the blood or brain of individuals with psychiatric diseases. Examples include aberrant DNA methylation of RELN, MB-COMT, HTR2A, ST6GALNAC1, AKT1, DNMT1, DTNBP1, NOS1, PPP3CC and 5-HTT in brain (31-36) and COMTD1, HTR1E, CD224, CD7, LAX1, MPG, MPO, PRF1, TXK, FAM63B and RELN in the blood (37-39) of patients with SCZ and/or BD, and AFF2, GABRB3, JMJD1C, KCNJ10, NLGN2, SNRPN, SNURF, PIK3C3 and UBE3A in the blood of autistic patients (40).

In the field of histone modifications, most of the studies have been related to SCZ and BD providing strong evidence for aberrant expression of several genes of histone proteins, including HIST1H2BC, HIST1H2BD, HIST1H2BH, HIST1H2BG, HIST1H4K and HIST2H2BE) in the blood cells of these patients and/or their first degree relatives (41, 42). Additionally, there are reports indicating i) an increase in the expression of H3-(methyl)arginine 17 in the prefrontal cortex associated with decreases in the expression of CRYM, MDH, OAT and CYTOC/CYC1 which are considered as metabolic genes (43), ii) aberrant expression of enzymes such as HDAC3 in the temporal cortex, which removes acetyl groups from the histone proteins decreasing gene expression (44), iii) an increased expression of HDAC1 in the frontal cortex, and an inverse association between the expression of HDAC1, HDAC3 or HDAC4 and GAD67 expression in SCZ patients (45).

Findings related to the roles of miRNA in mental disorders are also promising. Examples include, increased expression of miR15a and b, miR195, miR181b, miR107, exosomal miR29c and miR497 (46, 47) and decreased expression of miR24, miR26b, miR30e miR92 and miR346 (48) in the post-mortem brains of SCZ and/or BD patients. A decrease in the amount of circulating miR134 has been shown in the blood of BD patients in a manic phase, as well (49). In the blood of autistic patients increased expression of miR23a&b, miR132, miR146a and b, miR663, miR29b and miR103 and decreased expression of miR92a1/2, miR320, miR363, miR139-5p and miR219-5p were uncovered in miRNA microarray analysis (50, 51). However, the underlying origin of miRNA dysregulations as well as other epigenetic aberrations remained to be identified for preventive or therapeutic interventions.

Although hundreds of statistically significant genetic and/or epigenetic alterations have been found in major mental diseases, most of the epigenetic and even genetic alterations by themselves do not necessarily lead to disease phenotype. In fact, they should induce biologically significant

deleterious changes in the expression of coding genes or the structure of proteins to lead to the disease state. Considering the functional impacts of genetic/epigenetic alterations, at least at expression level, more than two dozen whole genome transcriptome analyses have taken place introducing hundreds of neuronal, synaptic and inflammatory genes with aberrant expression in the brain and/or blood of patients with psychiatric diseases (41, 52). Correlation of gene-specific or genome-wide epigenetic alterations with the expression of corresponding genes could identify epigenetic mechanisms of the altered functionality of more than a dozen genes related to synaptic transmission and axonal guidance (e.g. MB-COMT, HTR2A, 5-HTT, dopamine receptors, GAD1, RELN, DTNBP1, HTR1E, NOS1, PPP3CC, GRM5, PRIMA1, SHANK3) in major mental diseases, including autism (31-33, 36, 53-57).

It is important to note that, whereas some of these alterations might be tolerable in normal life, they may fail to adjust with specific conditions such as stressful life or exposure to contaminants and/or malnutrition. Hence, the dynamics of interactions between genome/epigenome and environment factors makes the issue more complex, a problem that in part can be assessed in a Petri dish using iPS cells of the affected or at risk individuals. Additionally, since epigenetic marks are mainly tissue-specific and blood epigenome cannot generally portray the brain epigenetic landscape, a number of studies undertook both brain and blood or saliva epigenetic analysis to identify peripheral epigenetic marks that represent the brain alterations. Dempster et al., found epigenetic aberrations of ST6GALNAC1 both in blood and brain of patients with psychosis (34). Others identified the same epigenetic alterations of MB-COMT, HTR2A and 5-HTT in brain and DNA extracted from the saliva of patients with SCZ or BD (36, 58, 59). Until now, these studies have not been undertaken in the same individuals using an adequate sample size, due to ethical issues associated with the brain biopsy in living individuals. Therefore, this important line of research remains to be

accomplished using novel approaches in coming years. Certainly, the detection of peripheral biomarkers mirroring the inaccessible brain tissues can revolutionize psychiatric evaluations with diagnostic, preventive or therapeutic applications.

### Psychiatric Drugs and Epigenetic Modifications

Several studies have shown that many current psychiatric drugs are, in fact, epigenetic modifiers. For example, valproate is a well-known HDAC inhibitor, and increases acetylated histone inducing gene expression, particularly in patients with BD (60). Lithium can also increase histone acetylation (61) and decrease global DNA methylation in patients who respond to this drug (62). Most of the antipsychotic drugs such as benzamides, clozapine, and lurasidone (63-65) and different classes of antidepressants (66-69) as well as ECT (70) are also epigenetic modifiers. There are also extensive efforts to identify novel epigenetic drugs that could target the affected genes/pathways for preventive or therapeutic remedies. While these efforts promise the development of novel drugs in near future, the lack of tissue specificity of most drugs remains a dilemma, not only in psychiatry, but also all branches of medicine which require to be addressed in coming years. Although new techniques such as TALEN and CRISPR are successfully used in laboratory experiments for mutational repairs and the delivery of epigenetic modifiers to specific genomic regions to modulate genes expression, these techniques are not likely to have clinical applications in the current decade.

### Conclusion

The fact that the same genetic mutations (1, 4, 7, 8) or epigenetic aberrations (epimutations), are linked to the pathogenesis of several psychiatric diseases is among the most interesting findings of recent GWAS and epigenetic analyses that support pleiotropic functions of these genes. Pleiotropy means that a specific gene may have several functions in different ages and/or

tissues. Hence, its malfunction may present various manifestations in different ages/tissues. For example, Tourette syndrome, ADHD, OCD, anxiety disorders and depression are closely linked together, so that the affected individual may show ADHD before tics in childhood, OCD in adolescence and anxiety and/or depression in adulthood (71). Therefore, a single drug may be appropriate for the adjustment of the activity of a pleiotropic gene improving all diverse phenotypes of that genetic/epigenetic disease in different ages or tissues.

Based on the studies reviewed here, the role of environment/nurture, including the uterine milieu might be more significant than genetic/nature in the genesis of most psychiatric diseases. While the normal epigenetic landscape is mostly established in the fetal period, followed by childhood and adulthood periods (72), several lines of evidence indicate that early life environmental impacts can alter epigenetic landscape with life-long effects. For examples, there are significant changes in DNA methylation profile of blood cells obtained from the umbilical cord of infants with arsenic exposure in utero (73). Furthermore, various factors such as nutritional imbalance, stress, smoking, drug abuse, birth weight and even seasonal changes in maternal diet during the periconceptional period may hamper the normal establishment of the epigenome (74-81). More importantly, many studies provided strong experimental evidence that the acquired epigenetic alterations can be retransferred to the next generations (82, 83) mimicking the inheritance of genetic diseases. Therefore, the shared environment of family members dictating the shared epigenetic portraits might be the origin of higher rate of familial psychiatric disorders misinterpreted as genetic diseases. Notably, even monozygotic twins with almost 100% genetic similarity only show ~45% concordance rate in SCZ that better fit with the idea of shared generational milieu, including uterine environment and maternal nutritional habits rather than genetic mutations.

The results of extensive genetic analyses during the last two decades, directing the

scientific community to rethink and revise the bolded roles of genetic factors in psychiatric and other complex diseases. Indeed, we are at the turning point of the age of genomics for reconsidering environmental factors that through epigenetic mechanisms may impact the brain development and functions at cellular or molecular levels. While epigenetic modifications may also balance the malfunction of genetic mutations, the ongoing deeper genetic/epigenetic analyses using the technology of next generation sequencing may help to recognize the combined genetic and epigenetic alterations leading to disease state with diagnostic, preventive and therapeutic applications in coming years.

### References

1. Lee SH, Ripke S, Neale BM, Faraone SV, Purcell SM, Perlis RH, et al. Genetic relationship between five psychiatric disorders estimated from genome-wide SNPs. *Nat Genet* 2013; 45(9): 984-94.
2. Doherty JL, O'Donovan MC, Owen MJ. Recent genomic advances in schizophrenia. *Clin Genet* 2012; 81(2): 103-9.
3. Hamshere ML, Walters JT, Smith R, Richards AL, Green E, Grozeva D, et al. Genome-wide significant associations in schizophrenia to ITIH3/4, CACNA1C and SDCCAG8, and extensive replication of associations reported by the Schizophrenia PGC. *Mol Psychiatry* 2013; 18(6): 708-12.
4. Green EK, Grozeva D, Jones I, Jones L, Kirov G, Caesar S, et al. The bipolar disorder risk allele at CACNA1C also confers risk of recurrent major depression and of schizophrenia. *Mol Psychiatry* 2010; 15(10): 1016-22.
5. Williams HJ, Craddock N, Russo G, Hamshere ML, Moskvina V, Dwyer S, et al. Most genome-wide significant susceptibility loci for schizophrenia and bipolar disorder reported to date cross-traditional diagnostic boundaries. *Hum Mol Genet* 2011; 20(2): 387-91.
6. Green EK, Hamshere M, Forty L, Gordon-Smith K, Fraser C, Russell E, et al. Replication of bipolar disorder susceptibility alleles and identification of two novel genome-wide significant associations in a

- new bipolar disorder case-control sample. *Mol Psychiatry* 2013; 18(12): 1302-7.
7. Lee KW, Woon PS, Teo YY, Sim K. Genome wide association studies (GWAS) and copy number variation (CNV) studies of the major psychoses: what have we learnt? *Neurosci Biobehav Rev* 2012; 36(1): 556-71.
  8. Purcell SM, Wray NR, Stone JL, Visscher PM, O'Donovan MC, Sullivan PF, et al. Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. *Nature* 2009; 460(7256): 748-52.
  9. Liu L, Sabo A, Neale BM, Nagaswamy U, Stevens C, Lim E, et al. Analysis of rare, exonic variation amongst subjects with autism spectrum disorders and population controls. *PLoS Genet* 2013; 9(4): e1003443.
  10. He X, Sanders SJ, Liu L, De RS, Lim ET, Sutcliffe JS, et al. Integrated model of de novo and inherited genetic variants yields greater power to identify risk genes. *PLoS Genet* 2013; 9(8): e1003671.
  11. Dickinson D, Straub RE, Trampush JW, Gao Y, Feng N, Xie B, et al. Differential effects of common variants in SCN2A on general cognitive ability, brain physiology, and messenger RNA expression in schizophrenia cases and control individuals. *JAMA Psychiatry* 2014; 71(6): 647-56.
  12. Wray NR, Pergadia ML, Blackwood DH, Penninx BW, Gordon SD, Nyholt DR, et al. Genome-wide association study of major depressive disorder: new results, meta-analysis, and lessons learned. *Mol Psychiatry* 2012; 17(1): 36-48.
  13. Ripke S, Wray NR, Lewis CM, Hamilton SP, Weissman MM, Breen G, et al. A mega-analysis of genome-wide association studies for major depressive disorder. *Mol Psychiatry* 2013; 18(4): 497-511.
  14. Stewart SE, Yu D, Scharf JM, Neale BM, Fagerness JA, Mathews CA, et al. Genome-wide association study of obsessive-compulsive disorder. *Mol Psychiatry* 2013; 18(7): 788-98.
  15. Sanders SJ, Murtha MT, Gupta AR, Murdoch JD, Raubeson MJ, Willsey AJ, et al. De novo mutations revealed by whole-exome sequencing are strongly associated with autism. *Nature* 2012; 485(7397): 237-41.
  16. Timms AE, Dorschner MO, Wechsler J, Choi KY, Kirkwood R, Girirajan S, et al. Support for the N-methyl-D-aspartate receptor hypofunction hypothesis of schizophrenia from exome sequencing in multiplex families. *JAMA Psychiatry* 2013; 70(6): 582-90.
  17. Yu TW, Chahrour MH, Coulter ME, Jiralerspong S, Okamura-Ikeda K, Ataman B, et al. Using whole-exome sequencing to identify inherited causes of autism. *Neuron* 2013; 77(2): 259-73.
  18. Cukier HN, Dueker ND, Slifer SH, Lee JM, Whitehead PL, Lalanne E, et al. Exome sequencing of extended families with autism reveals genes shared across neurodevelopmental and neuropsychiatric disorders. *Mol Autism* 2014; 5(1): 1.
  19. Ionita-Laza I, Xu B, Makarov V, Buxbaum JD, Roos JL, Gogos JA, et al. Scan statistic-based analysis of exome sequencing data identifies FAN1 at 15q13.3 as a susceptibility gene for schizophrenia and autism. *Proc Natl Acad Sci U S A* 2014; 111(1): 343-8.
  20. Takata A, Xu B, Ionita-Laza I, Roos JL, Gogos JA, Karayiorgou M. Loss-of-function variants in schizophrenia risk and SETD1A as a candidate susceptibility gene. *Neuron* 2014; 82(4): 773-80.
  21. Purcell SM, Moran JL, Fromer M, Ruderfer D, Solovieff N, Roussos P, et al. A polygenic burden of rare disruptive mutations in schizophrenia. *Nature* 2014; 506(7487): 185-90.
  22. Aberg K, Adkins DE, Bukszar J, Webb BT, Caroff SN, Miller DD, et al. Genomewide association study of movement-related adverse antipsychotic effects. *Biol Psychiatry* 2010; 67(3): 279-82.
  23. Adkins DE, Aberg K, McClay JL, Bukszar J, Zhao Z, Jia P, et al. Genomewide pharmacogenomic study of metabolic side effects to antipsychotic drugs. *Mol Psychiatry* 2011; 16(3): 321-32.
  24. McClay JL, Adkins DE, Aberg K, Stroup S, Perkins DO, Vladimirov VI, et al. Genomewide pharmacogenomic analysis of

- responseto treatment with antipsychotics. *Mol Psychiatry* 2011; 16(1): 76-85.
25. Niitsu T, Fabbri C, Bentini F, Serretti A. Pharmacogenetics in major depression: a comprehensive meta-analysis. *Prog Neuropsychopharmacol Biol Psychiatry* 2013; 45: 183-94.
  26. Hunter AM, Leuchter AF, Power RA, Muthen B, McGrath PJ, Lewis CM, et al. A genome-wide association study of a sustained pattern of antidepressant response. *J Psychiatr Res* 2013; 47(9): 1157-65.
  27. Ellsworth KA, Moon I, Eckloff BW, Fridley BL, Jenkins GD, Batzler A, et al. FKBP5 genetic variation: association with selective serotonin reuptake inhibitor treatment outcomes in major depressive disorder. *Pharmacogenet Genomics* 2013; 23(3): 156-66.
  28. Unternaehrer JJ, Daley GQ. Induced pluripotent stem cells for modelling human diseases. *Philos Trans R Soc Lond B Biol Sci* 2011; 366(1575): 2274-85.
  29. Mostafavi-Abdolmaleky H, Glatt ST. Epigenetics in psychiatry. In: Roach HI, Bronner F, Oreffo RO, editors. *Epigenetic aspects of chronic diseases*. London, UK: Springer Science & Business Media; 2011. p. 163-74.
  30. Mostafavi-Abdolmaleky H, Thiagalingam S. Pathogenic histone modifications in schizophrenia are targets for therapy. In: Peedicayil J, Grayson DR, Avramopoulos D, editors. *Epigenetics in psychiatry*. 1st ed. Philadelphia, PA: Elsevier; 2014. p. 241-51.
  31. Abdolmaleky HM, Cheng KH, Russo A, Smith CL, Faraone SV, Wilcox M, et al. Hypermethylation of the reelin (RELN) promoter in the brain of schizophrenic patients: a preliminary report. *Am J Med Genet B Neuropsychiatr Genet* 2005; 134B(1): 60-6.
  32. Abdolmaleky HM, Cheng KH, Faraone SV, Wilcox M, Glatt SJ, Gao F, et al. Hypomethylation of MB-COMT promoter is a major risk factor for schizophrenia and bipolar disorder. *Hum Mol Genet* 2006; 1(21): 3123-45.
  33. Abdolmaleky HM, Yaqubi S, Papageorgis P, Lambert AW, Ozturk S, Sivaraman V, et al. Epigenetic dysregulation of HTR2A in the brain of patients with schizophrenia and bipolar disorder. *Schizophr Res* 2011; 129(2-3): 183-90.
  34. Dempster EL, Pidsley R, Schalkwyk LC, Owens S, Georgiades A, Kane F, et al. Disease-associated epigenetic changes in monozygotic twins discordant for schizophrenia and bipolar disorder. *Hum Mol Genet* 2011; 20(24): 4786-96.
  35. Wockner LF, Noble EP, Lawford BR, Young RM, Morris CP, Whitehall VL, et al. Genome-wide DNA methylation analysis of human brain tissue from schizophrenia patients. *Transl Psychiatry* 2014; 4: e339.
  36. Abdolmaleky HM, Nohesara S, Ghadirivasfi M, Lambert AW, Ahmadkhaniha H, Ozturk S, et al. DNA hypermethylation of serotonin transporter gene promoter in drug naive patients with schizophrenia. *Schizophr Res* 2014; 152(2-3): 373-80.
  37. Nishioka M, Bundo M, Koike S, Takizawa R, Kakiuchi C, Araki T, et al. Comprehensive DNA methylation analysis of peripheral blood cells derived from patients with first-episode schizophrenia. *J Hum Genet* 2013; 58(2): 91-7.
  38. Liu J, Chen J, Ehrlich S, Walton E, White T, Perrone-Bizzozero N, et al. Methylation patterns in whole blood correlate with symptoms in schizophrenia patients. *Schizophr Bull* 2014; 40(4): 769-76.
  39. Aberg KA, McClay JL, Nerella S, Clark S, Kumar G, Chen W, et al. Methylome-wide association study of schizophrenia: identifying blood biomarker signatures of environmental insults. *JAMA Psychiatry* 2014; 71(3): 255-64.
  40. Wong CC, Meaburn EL, Ronald A, Price TS, Jeffries AR, Schalkwyk LC, et al. Methylomic analysis of monozygotic twins discordant for autism spectrum disorder and related behavioural traits. *Mol Psychiatry* 2014; 19(4): 495-503.
  41. Glatt SJ, Stone WS, Nossova N, Liew CC, Seidman LJ, Tsuang MT. Similarities and differences in peripheral blood gene-

- expression signatures of individuals with schizophrenia and their first-degree biological relatives. *Am J Med Genet B Neuropsychiatr Genet* 2011; 156B(8): 869-87.
42. Sanders AR, Goring HH, Duan J, Drigalenko EI, Moy W, Freda J, et al. Transcriptome study of differential expression in schizophrenia. *Hum Mol Genet* 2013; 22(24): 5001-14.
  43. Akbarian S, Ruehl MG, Bliven E, Luiz LA, Peranelli AC, Baker SP, et al. Chromatin alterations associated with down-regulated metabolic gene expression in the prefrontal cortex of subjects with schizophrenia. *Arch Gen Psychiatry* 2005; 62(8): 829-40.
  44. Aston C, Jiang L, Sokolov BP. Microarray analysis of postmortem temporal cortex from patients with schizophrenia. *J Neurosci Res* 2004; 77(6): 858-66.
  45. Sharma RP, Grayson DR, Gavin DP. Histone deacetylase 1 expression is increased in the prefrontal cortex of schizophrenia subjects: analysis of the National Brain Databank microarray collection. *Schizophr Res* 2008; 98(1-3): 111-7.
  46. Beveridge NJ, Gardiner E, Carroll AP, Tooney PA, Cairns MJ. Schizophrenia is associated with an increase in cortical microRNA biogenesis. *Mol Psychiatry* 2010; 15(12): 1176-89.
  47. Banigan MG, Kao PF, Kozubek JA, Winslow AR, Medina J, Costa J, et al. Differential expression of exosomal microRNAs in prefrontal cortices of schizophrenia and bipolar disorder patients. *PLoS One* 2013; 8(1): e48814.
  48. Perkins DO, Jeffries CD, Jarskog LF, Thomson JM, Woods K, Newman MA, et al. microRNA expression in the prefrontal cortex of individuals with schizophrenia and schizoaffective disorder. *Genome Biol* 2007; 8(2): R27.
  49. Rong H, Liu TB, Yang KJ, Yang HC, Wu DH, Liao CP, et al. MicroRNA-134 plasma levels before and after treatment for bipolar mania. *J Psychiatr Res* 2011; 45(1): 92-5.
  50. Talebizadeh Z, Butler MG, Theodoro MF. Feasibility and relevance of examining lymphoblastoid cell lines to study role of microRNAs in autism. *Autism Res* 2008; 1(4): 240-50.
  51. Sarachana T, Zhou R, Chen G, Manji HK, Hu VW. Investigation of post-transcriptional gene regulatory networks associated with autism spectrum disorders by microRNA expression profiling of lymphoblastoid cell lines. *Genome Med* 2010; 2(4): 23.
  52. Hwang Y, Kim J, Shin JY, Kim JI, Seo JS, Webster MJ, et al. Gene expression profiling by mRNA sequencing reveals increased expression of immune/inflammation-related genes in the hippocampus of individuals with schizophrenia. *Transl Psychiatry* 2013; 3: e321.
  53. Kordi-Tamandani DM, Sahranavard R, Torkamanzehi A. Analysis of association between dopamine receptor genes' methylation and their expression profile with the risk of schizophrenia. *Psychiatr Genet* 2013; 23(5): 183-7.
  54. Huang HS, Akbarian S. GAD mRNA expression and DNA methylation in prefrontal cortex of subjects with schizophrenia. *PLoS One* 2007; 2(8): e809.
  55. Xiao Y, Camarillo C, Ping Y, Arana TB, Zhao H, Thompson PM, et al. The DNA methylome and transcriptome of different brain regions in schizophrenia and bipolar disorder. *PLoS One* 2014; 9(4): e95875.
  56. Zhu L, Wang X, Li XL, Towers A, Cao X, Wang P, et al. Epigenetic dysregulation of SHANK3 in brain tissues from individuals with autism spectrum disorders. *Hum Mol Genet* 2014; 23(6): 1563-78.
  57. Zhubi A, Chen Y, Dong E, Cook EH, Guidotti A, Grayson DR. Increased binding of MeCP2 to the GAD1 and RELN promoters may be mediated by an enrichment of 5-hmC in autism spectrum disorder (ASD) cerebellum. *Transl Psychiatry* 2014; 4: e349.
  58. Nohesara S, Ghadirivasfi M, Mostafavi S, Eskandari MR, Ahmadkhaniha H, Thiagalingam S, et al. DNA hypomethylation of MB-COMT promoter in the DNA derived from saliva in



- schizophrenia and bipolar disorder. *J Psychiatr Res* 2011; 45(11): 1432-8.
59. Ghadirivasfi M, Nohesara S, Ahmadkhaniha HR, Eskandari MR, Mostafavi S, Thiagalingam S, et al. Hypomethylation of the serotonin receptor type-2A Gene (HTR2A) at T102C polymorphic site in DNA derived from the saliva of patients with schizophrenia and bipolar disorder. *Am J Med Genet B Neuropsychiatr Genet* 2011; 156B(5): 536-45.
  60. Sharma RP, Rosen C, Kartan S, Guidotti A, Costa E, Grayson DR, et al. Valproic acid and chromatin remodeling in schizophrenia and bipolar disorder: preliminary results from a clinical population. *Schizophr Res* 2006; 88(1-3): 227-31.
  61. Kwon B, Houpt TA. Phospho-acetylation of histone H3 in the amygdala after acute lithium chloride. *Brain Res* 2010; 1333: 36-47.
  62. Huzayyin AA, Andreazza AC, Turecki G, Cruceanu C, Rouleau GA, Alda M, et al. Decreased global methylation in patients with bipolar disorder who respond to lithium. *Int J Neuropsychopharmacol* 2014; 17(4): 561-9.
  63. Li J, Guo Y, Schroeder FA, Youngs RM, Schmidt TW, Ferris C, et al. Dopamine D2-like antagonists induce chromatin remodeling in striatal neurons through cyclic AMP-protein kinase A and NMDA receptor signaling. *J Neurochem* 2004; 90(5): 1117-31.
  64. Dong E, Nelson M, Grayson DR, Costa E, Guidotti A. Clozapine and sulpiride but not haloperidol or olanzapine activate brain DNA demethylation. *Proc Natl Acad Sci U S A* 2008; 105(36): 13614-9.
  65. Calabrese F, Luoni A, Guidotti G, Racagni G, Fumagalli F, Riva MA. Modulation of neuronal plasticity following chronic concomitant administration of the novel antipsychotic lurasidone with the mood stabilizer valproic acid. *Psychopharmacology (Berl)* 2013; 226(1): 101-12.
  66. Lee MG, Wynder C, Schmidt DM, McCafferty DG, Shiekhattar R. Histone H3 lysine 4 demethylation is a target of nonselective antidepressive medications. *Chem Biol* 2006; 13(6): 563-7.
  67. Perisic T, Zimmermann N, Kirmeier T, Asmus M, Tuorto F, Uhr M, et al. Valproate and amitriptyline exert common and divergent influences on global and gene promoter-specific chromatin modifications in rat primary astrocytes. *Neuropsychopharmacology* 2010; 35(3): 792-805.
  68. Tsankova NM, Bertone O, Renthal W, Kumar A, Neve RL, Nestler EJ. Sustained hippocampal chromatin regulation in a mouse model of depression and antidepressant action. *Nat Neurosci* 2006; 9(4): 519-25.
  69. Hunter RG, McCarthy KJ, Milne TA, Pfaff DW, McEwen BS. Regulation of hippocampal H3 histone methylation by acute and chronic stress. *Proc Natl Acad Sci U S A* 2009; 106(49): 20912-7.
  70. Tsankova NM, Kumar A, Nestler EJ. Histone modifications at gene promoter regions in rat hippocampus after acute and chronic electroconvulsive seizures. *J Neurosci* 2004; 24(24): 5603-10.
  71. Bloch MH, Leckman JF. Clinical course of Tourette syndrome. *J Psychosom Res* 2009; 67(6): 497-501.
  72. Numata S, Ye T, Hyde TM, Guitart-Navarro X, Tao R, Wininger M, et al. DNA methylation signatures in development and aging of the human prefrontal cortex. *Am J Hum Genet* 2012; 90(2): 260-72.
  73. Koestler DC, Avissar-Whiting M, Houseman EA, Karagas MR, Marsit CJ. Differential DNA methylation in umbilical cord blood of infants exposed to low levels of arsenic in utero. *Environ Health Perspect* 2013; 121(8): 971-7.
  74. Itzhak Y, Ergui I, Young JI. Long-term parental methamphetamine exposure of mice influences behavior and hippocampal DNA methylation of the offspring. *Mol Psychiatry* 2014.
  75. Joubert BR, Haberg SE, Nilsen RM, Wang X, Vollset SE, Murphy SK, et al. 450K epigenome-wide scan identifies differential DNA methylation in newborns

- related to maternal smoking during pregnancy. *Environ Health Perspect* 2012; 120(10): 1425-31.
76. Joubert BR, Haberg SE, Bell DA, Nilsen RM, Vollset SE, Middtun O, et al. Maternal smoking and DNA methylation in newborns: in utero effect or epigenetic inheritance? *Cancer Epidemiol Biomarkers Prev* 2014; 23(6): 1007-17.
  77. Teh AL, Pan H, Chen L, Ong ML, Dogra S, Wong J, et al. The effect of genotype and in utero environment on interindividual variation in neonate DNA methylomes. *Genome Res* 2014; 24(7): 1064-74.
  78. Engel SM, Joubert BR, Wu MC, Olshan AF, Haberg SE, Ueland PM, et al. Neonatal genome-wide methylation patterns in relation to birth weight in the Norwegian Mother and Child Cohort. *Am J Epidemiol* 2014; 179(7): 834-42.
  79. Waterland RA, Kellermayer R, Laritsky E, Rayco-Solon P, Harris RA, Travisano M, et al. Season of conception in rural gambia affects DNA methylation at putative human metastable epialleles. *PLoS Genet* 2010; 6(12): e1001252.
  80. Dominguez-Salas P, Moore SE, Cole D, da Costa KA, Cox SE, Dyer RA, et al. DNA methylation potential: dietary intake and blood concentrations of one-carbon metabolites and cofactors in rural African women. *Am J Clin Nutr* 2013; 97(6): 1217-27.
  81. Dominguez-Salas P, Moore SE, Baker MS, Bergen AW, Cox SE, Dyer RA, et al. Maternal nutrition at conception modulates DNA methylation of human metastable epialleles. *Nat Commun* 2014; 5: 3746.
  82. Waterland RA, Travisano M, Tahiliani KG, Rached MT, Mirza S. Methyl donor supplementation prevents transgenerational amplification of obesity. *Int J Obes (Lond)* 2008; 32(9): 1373-9.
  83. Manikkam M, Guerrero-Bosagna C, Tracey R, Haque MM, Skinner MK. Transgenerational actions of environmental compounds on reproductive disease and identification of epigenetic biomarkers of ancestral exposures. *PLoS One* 2012; 7(2): e31901.